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BIOEFFICACY OF PLANT EXTRACTS ON INHIBITION OF PYTHIUM MYRIOTYLUM

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ABSTRACT

Crude alcohol extract, 50% hydro-alcohol and aqueous extracts of 10 plants species belonging to 9 families were screened *in vitro* for antifungal activity against economically important phytopathogenic fungus, *Pythium myriotylum* which was isolated from infected ginger. Bioassays of the extracts were conducted by "Poisoned food technique" on agar plate culture with triplicates. Nine of ten (90%) plant species showed inhibitory activity against mycelial growth of the tested fungi. Among the 10 plants taken, *Jacaranda mimosifolia* showed best activity with 22.0% inhibition by 50% hydro-alcohol extract and 19.66% inhibition by its aqueous extract followed by *Moringa olifera* and *Lawsonia inermis* which exhibited 18.3% and 16.0% growth inhibition respectively. 13.0% and 12.6% inhibition was observed with *Terminallia arjuna* and *Polyalthia longifolia* respectively. All other selected plants exhibited inhibitory activity ranging from 5.3% to 10.0% against *Pythium myriotylum*. On the basis of these results, we conclude that the plants selected for this study can be regarded as a rich source of metabolites with significant antifungal activity, since 10mg/ml concentrations of crude extracts of these plants are showing significant activity. Partially purified fractions with high concentrations and active molecules may have enhanced activity against the test pathogen, as well as different species of *Pythium* and other fungi also.

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INTRODUCTION

Zingiber officinale Rosc. (Ginger), is a perennial rhizomatous herb belonging to the family Zingiberaceae (Hayden *et al.*, 2004) also an important commercial crop grown for its aromatic rhizomes which are used as a spice and medicine (Sharma *et al.*, 2010). It is a distinct family of aromatic tropical plants that yield spices, dyes, perfumes and medicines as well an important crop that earns a sizeable amount of foreign exchange for the country (Tarafdar and Saha, 2007). Ginger is a high return but also a high risk crop. Rhizome rot (also known as soft rot) is one of the most destructive diseases of ginger worldwide (Dohroo 2005). It reduces the potential yield of ginger to a great extent in the field, storage, and market and may cause more than 50 percent losses (Joshi and Sharma, 1980). 50–90% loss has been reported by Nirmal *et al.* (1992). Several practices have been used for the management of rhizome rot disease. Among them rhizome treatment with chemicals is one of the effective method which often provides some protection, against rhizome rot.

But it has residual effect and it is non-economical also. Hence biological control of this pathogen is a promising approach, seeing that it is comparatively benign towards the environment (Paulitz and Bélanger, 2001; Rattink, 1992). So the present study was conducted to investigate the inhibitory effect of crude alcohol, hydro-alcohol (50%) and aqueous extracts of plants given in the table no. 1 against *Pythium myriotylum*. The test pathogen was isolated from infected ginger rhizome.

MATERIALS AND METHODS

Collection, isolation and identification of the pathogen Diseased samples of ginger rhizomes were collected in sterilized polybags from various ginger farms in Jhadol, Udaipur, (Rajasthan) in the month of July –August. Plant samples were rinsed thoroughly under running tap water. Specimens were cut into 0.5-cm long segments, blotted dry on paper towels, and placed onto 2% water agar (Plaats-Niterink 1981). Cultures were incubated at room temperature (20-24°C) and observed daily for the emergence of fungal mycelium from the tissue. After 1–3 days, hyphal tips were removed from the colonies, transferred to V8 agar (Guo and Ko 1993), and identified according to the descriptions and key suggested by Plaats-Niterink (1981). Pure culture was maintained on PDA at 4 °C. Pure culture was also identified by Dr. Anila Doshi (Head,

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Department of Plant pathology, Rajasthan College of Agriculture Udaipur Rajasthan, India) as *Pythium myriotylum*.

Pathogenicity Test

5 days old culture of test pathogen growing on PDA plate was mixed in Sand-maize meal medium (9:1, 90gm of soil and 10gm of grinded maize). The mixture was kept for 10 days, then this inoculum was mixed with the top soil in the pot containing one month old plant of ginger. After 4 weeks of inoculum addition in the pot, disease severity was assayed by inoculating small pieces of leaves and rhizomes on WA. (Ghosh and Purkayastha 2003).

Preparation of Plant Extracts

Ten plants (Table No. 1) belonging to 9 different families were collected from the Botany Garden of University College of Science, Rajasthan college of Agriculture and from Fisheries Department, Udaipur. These botanicals were selected on the basis of presence of antimicrobial properties as given in the literature (Bobbarala et al., 2010, Pattnaik et al., 2012, Dileep et al., 2013, Garampalli and Rajkumar 2013). All the plants were identified by Dr. Maina, Head, BSI (Botanical Survey of India) Jodhpur, Rajasthan, India. Mature leaves of all the selected test plants were washed thoroughly with tap water, air dried in the shade on separate paper sheets then they were ground to a fine powder with the help of an electric blender. For extract preparation, 10gm of each powdered materials were added individually to 100ml of distilled water, 50% hydro-alcohol and 100% alcohol respectively and after 24 hours, the contents were filtered through four -fold muslin cloth followed by Whatman filter paper No.1 (Kekuda et al., 2010) and used for antifungal studies.

Table No. 1 List of Plants Screened for Antifungal Activity

S.No.	Name of the Plant	Vernacular Name	Family
1	Azadiracta indica	Neem	Meliaceae
2.	Aegle marmelos	Beel patrak	Rutaceae
3.	Cassia fistula	Amaltas	Fabaceae
4.	Jacaranda mimosifolia	Blue gulmohar	Bignoniaceae
5.	Lawsonia inermis	Mehandi	Lythraceae
6.	Moringa olifera	Sehjana	Moringaceae
7.	Murraya koenigii	Meetha neem	Rutaceae
8.	Polyalthia longifolia	Ashapal	Annonaceae
9.	Terminallia arjuna	Safeda	Combretaceae
10	Ziziphus jujuba	Jhadi ber	Rhamnaceae

Assay of in Vitro Antifungal Activity of Plant Extracts

In vitro antifungal efficacy of crude alcohol, 50% hydro-alcohol and aqueous, leaf extract against *Pythium myriotylum* was determined by Poisoned food technique (Groover and Moore 1962). 9 ml of PDA (Potato Dextrose Agar) media was mixed with 1ml (10mg/ml) of extract and sterilized in autoclave then poured into the sterilized Petri plates. A 5mm diameter fungal disc taken from actively growing 5 days-old culture of *Pythium myriotylum* on PDA, was placed in an inverted position in the centre of the Petri plates containing PDA amended with leaf extracts respectively. Plates containing medium with fungicide Mancozeb 0.2% (Indofil® mancozeb 75% WP) served as a positive control and plates with medium and 1ml of the solvents/water used to dissolve the extracts served as negative control. All plates were incubated at 28 °C and three replicates were maintained for each treatment. Radial growth of mycelium

was measured 5 days after inoculation. The results were compared with negative control. Experiment was repeated twice and mean of the readings were taken for calculations. The percent inhibition of the fungus in treatments was calculated using the following formula:

$$\text{Inhibition of mycelial growth (\%)} = (C-T/C) \times 100$$

Where 'C' is average diameter of fungal colony in control plates. 'T' is average diameter of fungal colony in poisoned plates (Gupta and Tripathi, 2011).

RESULTS

In the present study soft rot causing pathogen *Pythium myriotylum* was isolated from infected ginger rhizomes which were collected from Jhadol. The leaves of all the 10 selected plants were extracted in aqueous, 50% hydroalcohol and in 100% alcohol and their % extractive values are ranging from 1.0% to 21.15%. The highest % extractive value was found to be 21.15% followed by 17.55% which were from the hydro-alcohol and aqueous extracts of *Lawsonia inermis* respectively. The % extractive values of all the selected plant extracts are given in the table no. 2. Crude extracts of ten plants of the 9 species tested, showed 5.3% to 22.0% inhibitory activity against mycelial growth of *Pythium myriotylum* (Graph.no.1). Maximum inhibition of fungal growth was recorded by 50% hydro-alcohol extract of *Jacaranda mimosifolia* and it was found to be 22.0%. It was followed by 19.66% growth inhibition by aqueous extract of the same plant. 50% hydro-alcohol extract of *Moringa olifera* leaf also showed significant antifungal activity with 18.3% growth inhibition of *P. myriotylum*. Other plant extracts showed varying level of growth inhibition ranging from 5.3% to 16.0% (Table no.2).

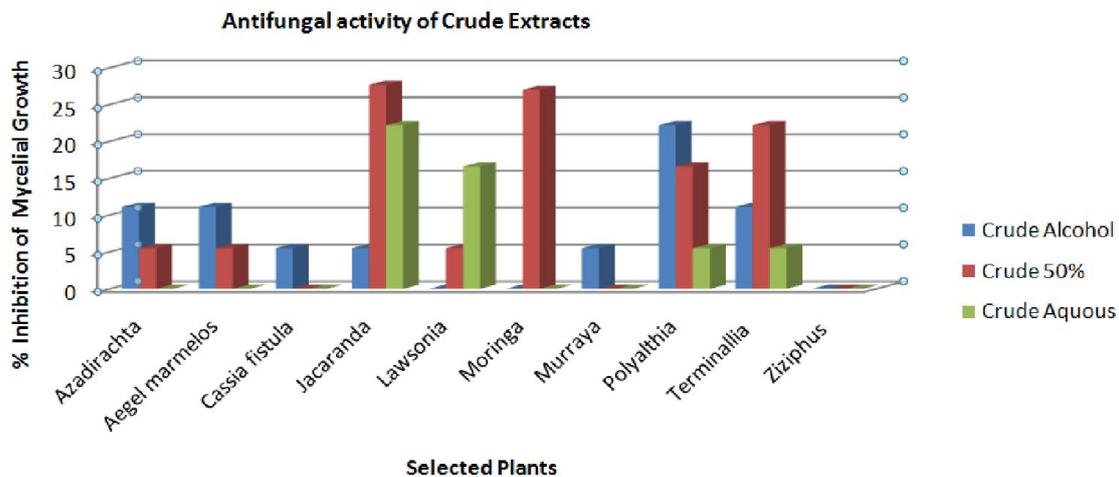
DISCUSSION

The genus *Pythium* is a complex genus containing over 200 described species that occupy a variety of terrestrial and aquatic ecological habitats (Dick, 2001). Perhaps the most economically important members of this genus are plant pathogens (Hendrix and campbell, 1973). Being very generalistic and unspecific in their host range, it is a major problem for a wide range of horticultural crops also. (Owen- Going 2002, Chaube and Pundhir 2005). *Pythium* species cause soft rot in ginger, Butler (1907) recorded the incidence of this disease for the first time from Surat (Gujarat, India). In India, at least six pathogenic species of *Pythium* have been reported to cause soft rot in ginger, and these include *P. myriotylum*, *P. aphanidermatum*, *P. deliense*, *P. perilium*, *P. vexans*, *P. ultimum* and *P. butleri* (Shahare and Asthana 1962, Haware and Joshi 1974, Dohroo 1987). *Pythium* species cause soft rot of ginger in Rajasthan, Himachal Pradesh, Orissa, Maharashtra, Tamil Nadu, Andhra Pradesh, and Sikkim (Singh et al., 2012). Lodha (2012) reported that *Pythium myriotylum* is the main species of *Pythium*, associated with the rhizome rot of ginger in Udaipur district. Around 90% of ginger produced in Rajasthan comes from Jhadol, a tribal-dominated block in Udaipur district. However, during the last 10 years both the area under cultivation and average productivity of ginger have shown a declining trend due to severe rot attack, a large number of farmers cultivated ginger in the region, but many gave up its cultivation owing to the frequent ginger rot disease that destroys the crops (ACCESS 2008).

Table No. 2 % Extractive Value of Extracts and % Inhibition of *Pythium myriotylum*

S.No.	Name of Plant	Extract Type	% Extractive value	% Inhibition \pm SD
1.	Azadiracta indica	Alcohol	2.65	12.59 \pm 0.6409
		50%hydro-alcohol	1.25	5.92 \pm 0.6409
		Aqueous	4.4	NA
2	Aegle marmelos	Alcohol	5.25	10.74 \pm 0.6409
		50%hydro-alcohol	6.25	5.92 \pm 0.6409
		Aqueous	17.3	NA
3	Cassia fistula	Alcohol	1.75	6.29 \pm 0.6409
		50%hydro-alcohol	1.00	NA
		Aqueous	2.25	NA
4	Jacarandas mimosifolia	Alcohol	8.50	5.92 \pm 0.6409
		50%hydro-alcohol	8.80	22.59 \pm 0.6409
		Aqueous	9.25	18.51 \pm 0.6409
5	Lawsonia inermis	Alcohol	9.95	NA
		50%hydro-alcohol	21.15	5.92 \pm 0.6409
		Aqueous	17.55	17.03 \pm 0.6409
6	Moringa olifera	Alcohol	6.50	NA
		50%hydro-alcohol	5.30	18.51 \pm 0.6409
		Aqueous	3.4	NA
7	Murraya koenigii	Alcohol	7.21	8.14 \pm 0.6409
		50%hydro-alcohol	4.30	NA
		Aqueous	3.85	NA
8.	Polyalthia longifolia	Alcohol	4.21	9.99 \pm 0.0058
		50%hydro-alcohol	2.82	11.85 \pm 0.6409
		Aqueous	2.73	6.29 \pm 0.6409
9.	Terminallia arjuna	Alcohol	3.25	5.92 \pm 0.6409
		50%hydro-alcohol	2.89	12.96 \pm 0.6409
		Aqueous	2.19	NA
10.	Zyzyphus zuzube	Alcohol	3.25	NA
		50%hydro-alcohol	2.89	NA
		Aqueous	2.19	NA
21	Mancozeb		100%	
22	Control C1		0%	
23	Control C2		0%	

NA: No Activity, C1: Negative control, C 2: Positive control

Graph 1. Efficacy of Various Extracts on % Inhibition of *Pythium myriotylum*

No single method is available to provide adequate control of the disease caused by *Pythium* (Babadoost 2004). Nowadays, synthetic pesticides are known to be the most effective method of the pest and disease control. However, they are not considered as a long-term solution due to the concerns associated with pesticides application such as problems of public health, environmental pollution, reduction in crop quality, toxic effect on non-target organisms and causing resistance in pest and disease agents, (Kagale 2004, Rai *et al.*, 2006, Rahhman *et al.*, 2010). WHO banned many agriculturally important pesticides due to wide range of toxicity against non target organisms including humans which are known to cause pollution problem (Barnard *et al.*, 1997). Besides this recent

studies have indicated that some isolates of *Pythium* are becoming less sensitive even to Metalaxyl (Daughtrey, 1998) which is a commonly used fungicide. This has necessitated search for alternatives for controlling the rhizome rot of ginger (Pandey *et al.*, 2010). In recent years, natural plant products as environmentally safe option have received attention for controlling phytopathogenic diseases. Many studies have shown that plant extracts effectively controlled various plant pathogens *in vitro* (Sankarasubramanian *et al.*, 2008, Mishra *et al.*, 2009, Yanar *et al.*, 2011, Talibi *et al.*, 2012). The fungicidal activity of some plant extracts in controlling different plant pathogens have been reported by several workers (Tewarri *et al.*, 1991, Amadioha 2000, Okigbo and Emoghene 2004, Okigbo and

Nmeka 2005). (Sagar *et al.*, 2007), (Haouala *et al.*, 2008) and (Suleiman and Emua 2009) also reported antifungal activity of plant extracts against *Pythium*. Although hundreds of plant species have been tested for antimicrobial properties, the vast majority of them have not been adequately evaluated (Balandrin *et al.*, 1985). The present study clearly demonstrates the significant inhibitory activity of various extracts of selected plants on rot causing pathogen *P. myriotylum* in *in vitro* condition. These results and the encouraging percentage of plants (90% in this research) with antifungal activity indicate that the plants selected can be regarded as rich sources of plants with antifungal activity. They could form the basis for further investigation of fractionation for finding active fractions. The present investigation was attempted to evaluate ten plants belonging to 9 different families of the plant kingdom to show the fact the plants are still a reservoir of many pharmaceuticals which can be isolated and used in plant disease management. It provide environmental friendly alternative to chemical fungicides for managing the pathogens.

Conclusion

From the results of the present study, it can be concluded that the crude extracts of selected plants are effective against the *Pythium myriotylum* and can be regarded as a rich source of metabolites with antifungal activity, the plant extracts which are showing inhibition for pathogen may have potential to be developed as potent fungicides in organic farming against rot causing pathogen. The plant world, the rich storehouse of natural chemicals could be exploited for the use as pesticides. Many species of higher plants have not been yet explored, much less surveyed for biologically active constituent and new sources of commercially valuable pesticides. This is mainly due to lack of information on the screening and evaluation of diverse plants for their antimicrobial potential. Systematic and scientific evaluation of plant derived bio-active molecules for using their potential for the effective management of fungal plant diseases to maintain high level of bio-safety and non-adverse effects on the environment will open up many revenues for the betterment of environment and mankind. The botanicals are cost effective, non hazardous, easily available and do not pollute the environment. Also, biologically active plant derived pesticides are expected to play a significant role in crop protection strategies. Exploitation of naturally available chemicals from plants, which retards the growth of disease causing pathogens, would be a more realistic and ecologically sound method for development of future commercial pesticides for crop protection strategies.

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REFERENCES

Access 2008, Rajasthan National Agriculture Innovation Project: Udaipur Region Ginger in Jhadol: Stemming the rot, ACCESS Development Services, 28, Hauz Khas Village, New Delhi – 16.

- Amadioha A C. 2000. Fungitoxic effects of some leaf extracts against *Rhizopus oryzae* causing tuber rot of potato Arch. Phytopathol. Pflanz, 0, 2000, 1-9.
- Babadoost M. 2004. Phytophthora blight: a serious threat to cucurbit industries. Available at: <http://www.apsnet.org/online/feature/cucurbit/links.asp>.
- Balandrin, M.F., Klocke, J.A., Wurtele, E.S. and Bollinger, W.H. (1985). Natural plant chemicals: Sources of Industrial and Medicinal materials. Science 228: 1154-1160.
- Barnard, C., Padgitt, M and Uri, N D. 1997. Pesticide use and its measurement. *International pest control*, 39, 1997, 161-164.
- Bobbarala varaprasad, Ammani K and Sunilbabu Kopula, 2010. Selected plant extracts as biocontrol agents against the management of Chilli (Capsicum annuum) disease, *Journal of Pharmacy research* 201, 3(12), 3143-3146.
- Butler E.J. An account of the genus *Pythium* and some Chytridiaceae. Mem. Dept. Agric. India, Bot. Ser. 1907;1(5):1-162.
- Chaube HS, Pundhir VS. (2005). Crop diseases and their management. Prentice-Hall of India. PP 702.
- Daugherty, M. 1998. Fungicide resistance in pathogens of greenhouse crops, p. 59-65. In: J. Hall and K. Robb (eds.). Proceedings for the 14th Conf. Insect and Disease Management on Ornamentals. Soc. Amer. Flor., Alex., Va.
- Dick, M.W., 2001. The peronosporomylates. In: the mycota VII part A. systematic Evolution (eds. D.J. McLaughlin, E.G., McLaughlin and P.A. Lenke), Springer verlag, Berlin. 39-72.
- Dileep, N., Junaid, S., Rakesh, K.N., Kekuda, P.T.R., Nawaz, N.A.S. 2013. Antifungal activity of leaf and pericarp extract of *Polyalthia longifolia* against pathogens causing rhizome rot of ginger. Science, *Technology and Arts Research Journal* 2(1): 56-59.
- Dohroo, N.P (2005) Bacterial Diseases of Ginger and their Control. In: Ravindran, P and Babu, K (Eds.) *Ginger: The Genus Zingiber*. Florida, USA: CRC Press pp 305-340
- Garampalli, H. Ravikumar, M.C and Rajkumar 2013. Archives of Phytopathology and Plant Protection, 2013, 1-7 pages <http://dx.doi.org/10.1080/03235408.2013.780350>.
- Ghosh R and Purkayastha RP (2003). Molecular diagnosis and induced systemic protection against rhizome rot disease of ginger caused by *Pythium aphanidermatum*, *Curr Sci* 85,1782
- Groover, R.K. and Moore, J.D. 1962. Toxicometric studies of fungicides against the brown rot organism *Sclerotinia fructivola* and *S. laxa*. *Phytopatho*, 52: 876-880.
- Guo L.Y. and W.H. Ko. 1993: Two widely Accessible Media for Growth and Reproduction of *Phytophthora* and *Pythium* Species: *Appl Environ Microbiol*. Jul 1993; 59(7): 2323-2325
- Gupta, S.K., Tripathi, S.C. 2011. Fungitoxic activity of *Solanum torvum* against *Fusarium sacchari*. *Plant Protection Science*, 47(3), 83-91.
- Haware M P and Joshi L K 1974. Studies on soft rot of ginger from Madhya Pradesh. *Indian Phytopath.* 27: 158-161.
- Hendrix, F.F. and Campbell, W., 1973. Annual Review of *Phytopathology*. 11: 78-98.
- Haouala R, Hawala S, ElA-yeb A, Khanfir R, Boughanmi N. 2008. Aqueous and organic extracts of *Trigonella foenum-graecum* L. inhibit the mycelia growth of fungi. *J Environ Sci* 20:1453-1457.
- H.K. Singh, R.C. Shakywar, Shyam Singh and Anil Kumar Singh, 2012. Evaluation of comparative efficacy of Native Isolate of *Trichoderma viride* against Rhizome rot Disease of Ginger. *Pl.Dis.Sci*. Vol 7(1) 2012 : 22 – 26.

- Hayden AL, Brigham LA, Gia ComelliGA, 2004. Aeroponic cultivation of Ginger (*Zingiber officinale*) Rhizome. ISHS Acta Horticulture, 2004:659(2):397-402.
- Joshi, L.K., and N.D. Sharma, 1980: Diseases of ginger and turmeric. In: Nair, M.K., Prem
- Kagale S, Marimuthu T, Thayumanavan B, Nandakumar R, Samiyappan R. 2004. *Physiol. Mol. Plant P.* 65:91-100.
- Kumar, T., Ravindran, P. N., and Sharma Y.R. (eds.) Ginger and Turmeric. CPCRI, Kasaragod, India, pp. 104–119.
- Kekuda, T.R.P., Kavya, R., Shrugashree, R.M., Suchithra, S.V. 2010. Screening of selected single and polyherbal ayurvedic medicines for antibacterial & antifungal activity. *Ancient Science of Life*, 29(3), 22-25.
- Lodha B.C.2012: Management of Rhizome Rot of Ginger: The Indian Science Congress Association Proc. 99th Indian Science Congress, Part II: Abstract of Symposium/Invited Lecture, SECTION OF PLANT SCIENCE (3-7 January 2012, Bhuvneshwar) Page no 287-288.
- Mishra AK, Mishra A, Kehri HK, Sharma B, Pandey AK. 2009. Inhibitory activity of Indian spice plant *Cinnamomum zeylanicum* extracts against *Alternaria solani* and *Curvularia lunata*, the pathogenic dematiaceous moulds. *Ann Clin Microbiol Antimicrob.* 8:9. doi: 10.1186/1476-0711-8-9.
- Nirmal K., Samsuddin K. and M.J.Ratnambal 1992 . *Plant, Cell tissue and Organ culture.* 29:71-74.
- Okigbo R N and Emoghene A O. Antifungal activity of leaf extracts of some plant species on *Mycosphaerella fijiensis* Morelet, the causal organism of black sigatoka disease of Banana (*Musa acuminata*) *KMITL Sci. J*, 4, 2004, 20-31.
- Okigbo R N and Nmeko I A. (2005)Control of yam tuber with leaf extracts of *Xylopiya aethiopica* and *Zingiber officinale*. *Afr. J. Biotechnol.* 4(8), 2005, 804 – 807.
- Owen-Going, T.N. 2002. Etiology and epidemiology of *Pythium* root rot in bell pepper (*Capsicum annuum*) in commercial-scale and small-scale hydroponic systems. M.Scthesis, U. of Guelph.
- Pandey, A.K., Awasthi, L.P., Srivastva, J.P., Sharma, N. K. (2010). Management of rhizome rot disease of ginger (*Zingiber officinale* Rose L.). *Journal of Phytology*, 2(9), 18-20.
- Paulitz, T.C. and R.R. Bélanger. 2001. Biological control in greenhouse systems. *Annu. Rev. Phytopathol.*, 39: 103-133.
- Plaats-Niterink, 1981 A.J. van der Plaats-Niterink, Monograph of the genus *Pythium*, *Stud. Mycol.* 21 (1981), pp. 1–244.
- Plaats-Niterink, A.J. 1981. Monograph of the genus *Pythium*. The Netherlands. Centraalbureau Voor Schimmelcultures 21: 1-242.
- Rai, M and Carpinella M. 2006, Naturally Occurring Bioactive compounds. Elsevier, Amsterdam 502 pp.
- Rattink, H. 1992. Targets for pathology research in protected crops. *Pestic. Sci.*, 36: 385-388.
- Rahman A and AM Hossain, 2010. *Eur Journal of Plant Pathol* 128: 211-219.
- Sankarasubramanian H, Duraiswamy S, Ramalingam R, Ebenezer EG, Seetharaman K. 2008. Use of plant extracts and biocontrol agents for the management of brown spot disease in rice. *Biocontrol.*53:555–567.
- Sharma, B.R., Dutta, S., Roy, S., Debnath A., Roy, M.D. 2010. The effect of soil physico-chemical properties on rhizome rot and wilt disease complex incidence of ginger under hill agro-climatic region of West Bengal. *Plant Pathology Journal*, 26(2), 198-202.
- Shahare K C and Asthana R P, 1962. Rhizome rot of ginger and its control. *Indian Phytopath.* 15: 77–78.
- Suleiman MN, Emua SA. 2009. Efficacy of four plant extracts in the control of root rot disease of cowpea (*Vigna unguiculata* [L.] Walp). *Afr J Biotechnol* 8(16):3806-3808.
- Sagar, S.D., Kulkarni, S., Hegde, Y.R. 2007. Management of rhizome rot of ginger by botanicals. *International Journal of Plant Science*, 2(2), 155-158.
- Talibi I, Askarne L, Boubaker H, Boudyach EH, Msanda F, Saadi B, Aoumar AAB. 2012. Antifungal activity of some Moroccan plants against *Geotrichum candidum*, the causal agent of post harvest citrus sour rot. *Crop Prot.* 35:41–46.
- Tarafdar, J., Saha, N. 2007. Correlation study on population dynamics of ginger soft rot inciting pathogens under different organic amendments, disease incidence and its survival in Darjeeling hill soils. Proceedings of the 13th ISTRC Symposium, 165-169.
- Yanar Y, Gokce A, Kadioglu I, Cam H, Whalon M. 2011. In vitro antifungal evaluation of various plant extracts against early blight disease (*Alternaria solani*) of potato. *Afr J Biotechnol.* 10:8291–8295.
